· Excise the DNA fragment from the Agarose gel with a clean, sharp scalpel.

(Minimize the size of the gel by removing extra agarose).

· Weigh the gel slice in a colourless tube. Add 3 volumes of Buffer QG to 1 volume of gel (100 mg~300µl).

· Incubate at 50°C for 10 mins. to dissolve agarose. Vortexing in between helps (37°C is fine).

· Add 1 gel volume of iso-propanol to the sample and mix (not for 500bp-4kb).

· Place a QIAquick spin column in a provided 2 ml collection tube.

· To bind DNA, apply the sample to the QIAquick column and centrifuge for 1min (maximum volume is 800µl, so do twice).

· Discard flow-through and place QIAquick column back in the same collection tube.

· Add 750µl Buffer PE to the column and centrifuge for 1min.

· Discard flow-through and centrifuge the column for an additional 1 min. at 13,000 rpm.

· Place the column into a clean 1.5ml eppendorf.

· To elute DNA, add 30µl Buffer EB or H_2O to the centre of the QIA membrane and centrifuge for 1 min.

· Can store at -20°C.